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ANTI-INFLAMMATORY COMPOSITIONS AND METHOD

This application claims the benefit of U.S. Provisional Application No. 60/112,020, which was converted pursuant to 37 C.F.R. § 1.53(c)(2)(i) from U.S. Patent Application No. 08/904,424, filed July 31, 1997, and of U.S. Provisional Application No. 60/054,453, filed August 1, 1997, all of which applications are incorporated herein by reference in their entireties.

10 **Field of the Invention**

The invention relates to compositions that modulate binding of a specific integrin molecule, $\alpha_9\beta_1$, to its receptor(s), to methods of treatment using such compounds, and to screening assays suitable for identifying additional modulatory compounds for use in such treatment methods. Pharmaceutical compositions which include such compounds are useful in treating inflammation and other disorders where modulation of $\alpha_9\beta_1$ -receptor interactions is desirable.

15 **References**

20 Hynes, R.O. (1987) *Cell* 48: 549-554.
Palmer, E.L., et al. (1993) *J. Cell Biol.* 123: 1289-1297.
Smith, et al. (1996) *J. Biol. Chem.* 271: 28485.
Yednock, T. A., et al. *J. Biol. Chem.*, 1995, 270: 28740-28750.
Yokosaki, et al. 1994, *J. Biol. Chem.* 269: 26691-26696.
Yokosaki, et al. 1996, *J. Biol. Chem.* 271: 24144-24150.

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Background of the Invention

The integrins are a group of glycoproteins that are present on a wide variety of cells, where they mediate cell-cell and cell-matrix adhesion via interactions with receptors present on cell membranes or in the extracellular matrix. Known receptors for

the various integrin family members include cell surface immunoglobulins, extracellular matrix proteins (laminin, collagen, fibronectin, tenascin), and cadherins.

All known members of the integrin family are composed of two subunits, termed alpha and beta. There are currently at least sixteen recognized alpha subunits and eight different beta-subunits; integrins containing the β_1 form of the beta subunit are known as the " β_1 integrin family." Members of this family are expressed by a diverse distribution of tissues and exhibit specific binding specificities. Thus, $\alpha_1\beta_1$ integrin is expressed by T-lymphocytes and fibroblasts and binds to collagen and laminin; in contrast, $\alpha_4\beta_1$ integrin (VLA-4) is expressed by several types of hematopoietic cell and binds to 10 VCAM-1, fibronectin and madCAM. It is therefore the alpha subunit that apparently confers receptor binding specificity to the protein.

A relatively new member of the β_1 integrin family, $\alpha_9\beta_1$ (also referred to herein as "alpha-9 integrin") has been shown to bind to tenascin and osteopontin, both of which are components of the extracellular matrix which are induced at sites of inflammation 15 (Yokosaki; Smith). When sequences of the various alpha subunits were compared, alpha-9 integrin was shown to have the closest sequence identity to the alpha-4 subunit; however this represents only 39% sequence identity (Palmer). Moreover, the two subunits have different cell and tissue distributions. While $\alpha_9\beta_1$ is expressed on airway smooth muscle cells, and non-intestinal epithelial cells (Palmer), and diffusely on 20 hepatocytes and basal keratinocytes (Yokosaki, 1994), $\alpha_4\beta_1$ integrin is present mainly on hematopoietic cells.

Heretofore, there has been no definitive determination of an *in vivo* function for $\alpha_9\beta_1$ integrin, nor has a physiological consequence of disruption of $\alpha_9\beta_1$ - receptor interactions been identified, despite its presence in several tissues, as described above. 25 Nor, despite its association with osteopontin and tenascin, has there been any reason to suspect that alpha-9 integrin might play a role in inflammatory disorders, since the $\alpha_9\beta_1$ molecule had not been associated with any of the hematopoietic cells commonly associated with this disorder.

In studies carried out in support of the present invention, it is now found that 30 $\alpha_9\beta_1$ is present on neutrophils, a class of phagocytic cells which play an important role in

inflammation. In humans, these cells are notable for their relative lack of alpha-4/beta-1 integrin. Therefore, the present invention provides basis for involvement of $\alpha_9\beta_1$ in acute inflammatory responses.

Further differences among the β_1 -integrins are associated with their binding specificities or endogenous ligands. While they all bind one or more proteins or proteoglycans that form the extracellular matrix, each integrin family member exhibits a distinct molecular specificity which may dictate, in part, its physiological specificity. Thus, while alpha-4/beta-1 integrin is known to bind fibronectin and VCAM-1, alpha-9 integrin has been characterized as binding the matrix proteins osteopontin and tenascin 10 (Yokosaki, 1994; Smith, 1996). According to a further discovery related to the present invention, alpha-9 integrin also binds VCAM-1, though, as discussed below, it is likely that such binding occurs at a site that distinct from the alpha-4 binding site.

The present invention therefore provides basis for new therapeutic regimens directed at modulating alpha-9 integrin binding to its ligand(s), and in particular, those 15 ligands which are involved in the inflammatory response. In addition, it is a further discovery of the present invention that many of the compounds or drugs that modulate (inhibit or enhance) alpha-4/beta-1 integrin binding also modulate alpha-9 integrin binding. This discovery therefore provides new pharmaceutical compositions and methods of treatment for modulating alpha-9 integrin binding, as well as screening 20 methods for identifying new alpha-9 integrin modulatory compounds.

Summary of the Invention

The invention is directed to pharmaceutical compositions and methods of treatment for disorders that involve binding of alpha-9 integrin, as well as screening 25 assays that are useful in identifying compounds for use in such compositions and methods. More particularly, the invention is directed to inflammatory conditions, particularly those that involve increased adhesion macrophages or neutrophils, which, according to a discovery of the present invention, are now known to carry alpha-9 integrin in their membranes and to exhibit increased expression of alpha-9 integrin in 30 response to stimulation by a known activator molecule, fMLP, as described herein.

A number of inflammatory disorders are therefore susceptible to treatment in accordance with the present invention, including but not limited to airway hyper-responsiveness and occlusion that occur in conjunction with chronic asthma, smooth muscle cell proliferation in atherosclerosis, vascular occlusion following angioplasty, 5 fibrosis and glomerular scarring as a result of renal disease, aortic stenosis, hypertrophy of synovial membranes in rheumatoid arthritis, and inflammation and scarring that occur with the progression of ulcerative colitis, and Crohn's disease.

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10 ~~In preferred embodiments, pharmaceutical compositions and methods of treatment of the invention employ alpha-9 antagonist compounds that inhibit binding between alpha-9 integrin and an alpha-9 integrin ligand. Preferred ligands in this regard include any ligand found to specifically bind to alpha-9 integrin, as exemplified by osteopontin, tenascin, and VCAM-1. Due to its association with inflammatory reactions, V-CAM-1 is particularly preferred for a test compound.~~

15 In one embodiment, pharmaceutical compositions and treatment methods of the invention contain an alpha-9 integrin antagonist compound that exhibits a potency in inhibiting binding between alpha-9 integrin and an alpha-9 integrin ligand that is at least as high as 1/1000, and preferably at least as high as 1/100 of an inhibitory potency exhibited by a compound selected from the group consisting of nine reference compounds: N-(toluene-4-sulfonyl)-L-prolyl-L-4(4-methylpiperazin-1-ylcarbonyloxy)phenylalanine, N-(toluene-4-sulfonyl)-L-prolyl-L-4(N,N-dimethylcarbamyl)phenylalanine, N-(1-methylpyrazole-4-sulfonyl)-L-prolyl-L-4(N,N-dimethylcarbamyl)phenylalanine, N-(toluene-4-sulfonyl)-L-(1,1-dioxo-5,5-dimethyl)thiaprolyl-L-4-(N,N-dimethylcarbamyl)phenylalanine, N-(toluene-4-sulfonyl)-N-methyl-L-alaninyl-L-4-(N,N-dimethylcarbamyl)phenylalanine, N-20 (toluene-4-sulfonyl)-L-[1,1-dioxo]thiamorpholin-3-carbonyl]-L-4-(N,N-dimethylcarbamyl)phenylalanine, N-(N-p-toluenesulfonyl)prolyl-4-(piperazinoyloxy)phenylalanine, N-(N-p-toluenesulfonyl)sarcosyl-4-(N,N-dimethylcarbamyl)phenylalanine, and N-(toluene-4-sulfonyl)-L-(5,5-dimethyl)thiaprolyl-L-4-[3-(N,N-dimethyl)propoxy]phenylalanine.

The foregoing group of compounds are exemplary in nature, having been chosen for their relatively high potency in inhibiting alpha-9 integrin binding to an exemplary ligand, tenascin. The foregoing compounds also illustrate another aspect of the invention – that a rich source of candidate compounds for use in the pharmaceutical compositions and methods of treatment described herein is compounds known to inhibit binding or activity of alpha-4/beta-1 integrin (VLA-4). The foregoing 9 reference standard compounds can also be used in the pharmaceutical compositions and methods of treatment described above.

In another embodiment, pharmaceutical compositions and methods of treatment will employ alpha-9 integrin antagonists that have a K_i or IC_{50} less than about 100 μM , as determined in an assay which measures inhibition of binding between alpha-9 integrin and an alpha-9 integrin ligand.

In another related embodiment, compounds useful in the pharmaceutical compositions and methods of treatment of the invention have the formula:

R⁵ is -(CH₂)_x-Ar-R^{5'} where R^{5'} is selected from the group consisting of -O-Z-NR⁸R^{8'} and -O-Z-R¹² wherein R⁸ and R^{8'} are independently selected from the group consisting of hydrogen, alkyl, substituted alkyl, cycloalkyl, substituted cycloalkyl, heterocyclic, substituted heterocyclic, and where R⁸ and R^{8'} are joined to form a heterocycle or a substituted heterocycle, R¹² is selected from the group consisting of heterocycle and substituted heterocycle, and Z is selected from the group consisting of -C(O)- and

5 -SO₂-,

Ar is aryl, heteroaryl, substituted aryl or substituted heteroaryl,

10 x is an integer of from 1 to 4;

Q is -C(X)NR⁷ - wherein R⁷ is selected from the group consisting of hydrogen and alkyl; and X is selected from the group consisting of oxygen and sulfur, and pharmaceutically acceptable salts thereof.

In still another related embodiment, the invention includes pharmaceutical

15 compositions and methods which employ a small molecule compound. The compound is selected for its ability to inhibit binding between alpha-9 integrin and an alpha-9 integrin ligand, as evidenced by exhibiting a potency in an alpha-9 integrin-alpha-9 integrin ligand binding assay that is at least 1/1000 as high as a potency of a compound selected from the reference compound group listed above. In a related embodiment, such a
20 compound is also an inhibitor of inhibitor of alpha-4/beta-1 integrin binding to VCAM-1, as evidenced by its ability to inhibit such binding with a potency that is at least 1/1000 as high as a potency exhibited by a compound selected from the reference standard group listed above. All pharmaceutical compositions and methods of treatment employ pharmaceutically effective dosages and are delivered in an excipient and manner
25 appropriate to the particular treatment regimen selected by the practitioner.

According to a related aspect, the invention includes a method of screening for therapeutic compounds effective in treating conditions characterized by involvement of alpha-9 integrin, and in particular, inflammatory conditions, such as those listed above.

30 The method includes adding test compound to an assay system which measures an amount of alpha-9 integrin binding to an alpha-9 integrin ligand, and selecting the test

compound as an effective therapeutic drug candidate, if said compound exhibits a binding inhibitory activity that is at least 1/1000 as potent as an activity exhibited by a compound selected from the group of reference standard compounds listed above. Candidate compounds selected in this mode are further tested for safety and toxicity, according to 5 methods well known in the art, prior to use in the pharmaceutical compositions and methods of treatment described herein.

Selection of test compounds for testing in the screening method is well within the skill of the practitioner in view of the wealth of combinatorial libraries now commercially available or available through the scientific literature. Nonetheless, in 10 accordance with the present invention, particularly preferred test compounds are those known to exhibit activity in modulating, particularly inhibiting, binding between alpha-4/beta-1 integrin and any of its ligands, but particularly VCAM-1.

According to a preferred embodiment, a compound is selected by the assay if it exhibits an inhibitory potency that is at least as 1/1000 as high as an inhibitory potency exhibited 15 by a compound selected from the reference standard group listed above. According to a related embodiment, preferred compounds are selected from a group having the formula: $R^1-SO_2-NR_2-CHR^3-Q-CHR^5-CO_2H$, where the substituent groups and moieties are defined as described above. These compounds are also described in co-owned parent applications U.S. Patent application 08/904,424, filed July 31, 1997, and U.S. Provisional 20 Application No. 60/054,453, filed August 1, 1997, which are incorporated herein by reference.

These and other objects and features of the invention will become more fully apparent when the following detailed description of the invention is read in conjunction with the accompanying drawings.

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Brief Description of the Figures

FIG. 1 shows a bar graph that illustrates expression of alpha-9 integrin on human neutrophils; and

FIG. 2 shows a bar graph illustrating selective enhancement of expression of alpha-9 integrin ("anti- α 9 integrin) as compared to control ("isotype control") on human neutrophils following activation by fMLP (right-hand bars).

5 Detailed Description of the Invention

I. Definitions

This section provides definitions of certain of the terms used herein. Unless specifically defined, all other scientific and technical terms used have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. A convenient reference for purposes the present invention is Stedman's Medical Dictionary 24th Edition (Williams and Wilkins, Baltimore)

α₉β₁
The term "alpha-9 integrin" refers to a heterodimeric protein member of the β_1 integrin family which is also referred to as $\alpha_9\beta_1$.

The term "alpha-9 integrin ligand" refers to molecules to which alpha-9 integrin binds *in vitro* or preferably *in vivo*. Preferably, such binding occurs with a binding affinity or potency in the range of at least about 10⁻⁴ M and typically between about 10⁻⁵ to 10⁻⁸ M. The term also refers to fragments of such compounds that possess such binding affinity characteristics. Exemplary alpha-9 integrin ligands include, but are not limited to tenascin, osteopontin, and VCAM-1.

The term "binding affinity" as used herein refers to the relative strength with which two or more molecules bind together. In its usage herein, the term is typically expressed in terms of the molar amount of compound necessary to observe a desired effect, such as ½ maximal binding or response (EC₅₀ or K_d), or inhibition of binding (IC₅₀ or K_i), but may also be used in a relative sense to express an amount of compound required to observe minimal saturation of a ligand, such as in the integrin saturation assays described herein.

The term "potency" is generally used to refer to relative affinities or efficacies; a compound has a "higher potency" than another if it is more effective than a reference compound when the two are compared at the same molar concentration, or if it produces the same effect at a lower concentration. The term may also be used to compare amounts

of different compound needed to observe an arbitrary effect. By way of example, a test compound is said exhibits “a potency that is at least as high as 1/1000 of a reference standard potency, if it produces the same effect as the reference standard at no more than 1000 times the molar concentration required for the reference standard. Therefore, if the 5 reference standard produces a given effect at a concentration of 1 μ M, a test compound would be at least 1/1000 as potent if it is capable of producing the same effect at any concentration less than 1000 μ M (1 mM).

The term “pharmaceutical composition” refers to a pharmaceutically active preparation of drug or biological which is prepared in a pharmaceutical excipient, such as 10 buffered saline or a physiological buffer appropriate for administration to a subject. Appropriate excipients, including but not limited to diluents, fillers and the like are formulated based on the anticipated mode of administration and are readily determined by persons skilled in the art.

The term “small molecule” generally refers to an organic compound having a 15 molecular weight that is less than about 2000 and preferably less than about 1000; small molecules may include short peptides and peptidomimetics.

The term “alpha-4/beta-1 integrin” refers to a heterodimeric protein which is also referred to as $\alpha_4\beta_1$ and as VLA-4.

The term “alpha-4/beta-1 integrin ligand” refers to molecules to which alpha-20 4/beta-1 integrin binds *in vitro* or preferably *in vivo*. Preferably, such binding occurs with a binding affinity in the range of at least about 10^{-4} M and typically between about 10^{-5} to 10^{-8} M. The term also refers to fragments of such compounds that possess such binding affinity characteristics. Exemplary alpha-4/beta-1 integrin ligands include, but are not limited to fibronectin (HEPII and CS1 domains), VCAM-1, osteopontin, and madCAM1. 25 Generally, such ligands include a peptide binding site having the sequence EILDV.

The term “condition associated with binding of alpha-9 integrin to an alpha-9 integrin ligand” describes conditions having attributes consistent with the presence of increased or reduced alpha-9 integrin bound to an endogenous alpha-9 integrin ligand as compared to normal. An example of increased alpha-9 integrin binding is increased 30 adhesion of neutrophils to osteopontin or tenascin, or another alpha-9 ligand during

inflammation. In view of studies carried out in support of the present invention and described herein, alpha-9 integrin is likely to be involved in such increased adherence. Thus inflammation is considered a condition associated with binding of alpha-9 integrin.

An “alpha-9 integrin modulatory compound” or an “alpha-9 integrin regulatory compound” is a compound, preferably but not necessarily a small molecule, which, when added to a mixture which contains alpha-9 integrin and an alpha-9 integrin ligand, affects the binding between the integrin molecule and the ligand – for example, by producing an increase or decrease of such binding. By way of example, a modulatory compound which inhibits or reduces alpha-9 integrin binding to tenascin is referred to as an alpha-9 antagonist.

An effect or response is “significantly different,” “significantly higher” or “significantly lower” if, when compared to an appropriate control, the test response shows a statistically significant change, when analyzed by an appropriate statistical method. In general, when it is stated that a response or effect is increased or decreased, it can be inferred that the observed increase or decrease is statistically significant or is expected to be statistically significant when subjected to appropriate experimental analysis.

Common amino acids are referred to by their one- or three-letter abbreviations as follows: alanine (A, Ala), cysteine (C, Cys), aspartic acid (D, Asp), glutamic acid (E, Glu), phenylalanine (F, Phe), glycine (G, Gly), histidine (H, His), isoleucine (I, Ile), lysine (K, Lys), leucine (L, Leu), methionine (M, Met), asparagine (N, Asn), proline (P, Pro), glutamine (Q, Gln), arginine (R, Arg), serine (S, Ser), threonine (T, Thr), valine (V, Val), tryptophan (W, Trp), tyrosine (Y, Tyr).

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II. Alpha-9 Integrin

This section provides further background information about alpha-9 integrin, including means for distinguishing it from other integrins. Such distinguishing means are important in (i) setting up assays capable of measuring binding or blockade of binding between $\alpha_9\beta_1$ integrin and its ligand(s), and (ii) identifying compounds that are preferably

small molecule antagonists capable of blocking interactions between $\alpha_9\beta_1$ integrin and its ligands(s).

A. Physical Characteristics

As mentioned above, $\alpha_9\beta_1$ integrin is a heterodimeric protein consisting of an alpha-subunit, termed α_9 (or "alpha-9"), and a beta-subunit, generally β_1 (or "beta-1"). The two subunits bind to one another noncovalently, and each consists of a relatively short carboxy terminal intracellular domain which contains the highly conserved sequence GFF(R/K)R, a single transmembrane domain and a relatively large amino terminus extracellular domain which generally projects on the surface of cells.

The deduced amino acid sequence of the alpha-9 subunit has been determined by cloning (Palmer, 1993; GENBANK Accession No. L24158) The human alpha-9 subunit is a protein of 1006 amino acids. Studies comparing the various forms of alpha subunits have revealed that the alpha-9 subunit exhibits only 39% sequence identity with the alpha-4 integrin subunit.

B. Tissue Localization and Binding Selectivity of alpha-9 Integrin
 $\alpha_9\beta_1$ integrin is expressed by a number of different cell types, as mentioned above. For example, $\alpha_9\beta_1$ is found on airway smooth muscle cells, and non-intestinal epithelial cells, as well as a teratoma cell line (Palmer), and diffusely on hepatocytes and basal keratinocytes (Yokosaki, 1994). Heretofore, alpha-9 has not been shown to be present on any of the hematopoietic cells.

FIG. 1 shows results of experiments carried out in support of the present invention that show that, in addition to the previously known tissue distribution described above, alpha-9 integrin is also expressed by human neutrophils. In this experiment, alpha-9 subunit, alpha-4 subunit and beta-1 subunit were measured after reacting human neutrophils with fluorescently labeled antibodies specifically reactive with each of the foregoing subunits. This shows that, surprisingly, human neutrophils express alpha-9 subunit along with beta-1 subunit, and confirms that they express very little, if any alpha-4 subunit.

Neutrophils are phagocytic blood cells that are involved in a number of inflammatory conditions, particularly acute inflammation, as described below. These cells were previously distinguished by their lack of alpha-4/beta-1 integrin, which is expressed by all, or nearly all other circulating leucocytes.

5 Further experiments in support of the invention indicated that alpha-9 integrin is likely involved in inflammatory responses involving neutrophils. Formyl-Met-Leu-Phe (fMLP) is an activation factor that is involved in inflammation. FIG. 2 is a bar graph that shows that alpha-9 integrin expression is significantly and selectively increased following activation by fMLP (right-hand bars), as compared to a control idiotype-specific marker. This increased expression is consistent with alpha-9 involvement in 10 activation of neutrophils during inflammation.

Studies on the binding selectivity of alpha-9 integrin have revealed that the molecule binds to tenascin at a “fibrinogen-like” type-III repeat (termed “TNfn3”). Although this region of tenascin contains the characteristic “RGD” (Arg-Gly-Asp) 15 peptide binding site to which other integrins ($\alpha_8\beta_1$, $\alpha_v\beta_3$, and $\alpha_v\beta_6$) preferentially bind, this is apparently not the site bound by alpha-9 integrin (Yokosaki, 1994). Rather, $\alpha_9\beta_1$ integrin binds preferentially to the B-C loop which contains the peptide sequence AEIDGIEL, and a peptide containing this sequence has been shown to disrupt binding between cells expressing alpha-9 integrin and TNFn3 (Yokosaki, 1998).

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III. Alpha-9 Integrin Modulatory Compounds

This section provides guidance for identifying compounds for use as alpha-9 integrin modulatory compounds suitable for use in pharmaceutical compositions and treatment methods in accordance with the present invention. Specifically, in studies 25 carried out in support of the present invention it has been found that, despite their sequence and ligand binding site dissimilarities noted above, alpha-4/beta-1 integrin and alpha-9 integrin apparently share similar binding sites for small molecules, and that binding to such sites serves to similarly modulate their abilities to bind to endogenous ligands.

Therefore, according to a preferred aspect of the present invention, compounds that modulate alpha-4/beta-1 integrin binding to its ligands are likely to also have activity in alpha-9 integrin assays. Accordingly, as described below, a rich source of compounds for testing in specific alpha-9 integrin modulatory assays consists of small molecules, 5 including peptides and peptidomimetics, that have been characterized as alpha-4/beta-1 integrin agonists or antagonists. Further candidate compounds are provided by a variety of libraries, including, without limitation, combinatorial libraries, fermentation broths and lysates, phage libraries, and the like, such as are well known in the art and/or commercially available for screening, as discussed in more detail in Part B, below. Such 10 libraries of compounds, in addition to the alpha-4/beta-1 modulatory compounds mentioned above and any other compounds can be conveniently screened in assay formats known in the art with reference to the exemplary assays described herein. Such assays can be further modified to accommodate high throughput screening of compounds according to methods known in the art.

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A. Alpha-4/beta-1 integrin Agonists and Antagonists

In contrast to alpha-9 integrin, which has only recently been characterized, alpha-4/beta-1 integrin has been widely studied and has been the focus of numerous drug development programs. Alpha-4/beta-1 integrin is expressed by most forms of 20 hematopoietic cells, with the exception of neutrophils (e.g., $\alpha_4\beta_1$ is expressed by T- and B-lymphocytes, monocytes and certain antigen presenting cells). Alpha-4/beta-1 integrin binds endogenous ligands including fibronectin, mucosal addressin (MadCAM-1), vascular cell adhesion molecule-1 (VCAM-1; Cd106) and osteopontin. In particular, its interaction with VCAM-1, which is induced in the vascular epithelium during acute 25 inflammatory responses, its presence on leukocytes, and its recognized involvement in the enhanced adhesion of such leukocytes at sites of inflammation have made $\alpha_4\beta_1$ a target for compounds in development for treatment of a variety of inflammatory diseases, including rheumatoid arthritis, heart disease and ulcerative colitis, among others.

Studies carried out in support of the present invention have revealed that 30 compounds which modulate binding of alpha-4/beta-1 integrin to any of its ligands,

including VCAM-1, are generally also good candidates for modulating binding of alpha-9 integrin to its ligand(s). More specifically, as described in section IV, below, specific antagonists of alpha-4/beta-1 integrin also block alpha-9 integrin binding to tenascin.

This discovery therefore provides a wealth of candidate compounds for use in

5 pharmaceutical compositions and methods of the present invention. For example, co-owned, concurrently-filed related application USSN _____, (PCT _____) incorporated herein by reference, which claims priority to the same initial U.S. Patent Applications, U.S. Patent Application No. 08/904,424, filed July 31, 1997, and U.S. Provisional Application No. 60/054,453, filed August 1, 1997, which are incorporated 10 herein by reference in their entireties, describes a series of carbamyloxy compounds which have been characterized as inhibitors of alpha-4/beta-1 integrin binding to its ligands (*i.e.*, $\alpha_4\beta_1$ antagonists). Testing and use of several exemplary compounds of this series are discussed below. Methods for making such compounds are detailed in Example 4 herein.

15 Similarly, co-owned U.S. patent applications USSN 08/904,415, USSN 08/903,585, USSN 08/904,423, USSN 08/920,353, USSN 08/904,417, USSN 08/920,394 and USSN 08/904,416, all filed on July 31, 1997, describe additional, structurally distinct alpha-4/beta-1 integrin antagonists which inhibit binding of alpha-4/beta-1 integrin to its ligand(s). The foregoing applications are hereby incorporated herein by reference for 20 their teachings of such compounds which, in accordance with the present invention, are also candidates for use in the pharmaceutical compositions and methods of treatment that are the subjects of the present invention. In view of the data presented below, it is anticipated that many of these compounds will exhibit approximately equipotent inhibitory activities in inhibiting alpha-9 integrin as in inhibiting alpha-4/beta-1 integrin.

25 **B. Sources of Test Compounds**

Additional sources of candidate alpha-9 integrin modulatory compounds are therefore apparent, in view of the discovery that at least a significant subset of alpha-4/beta-1 (VLA-4) inhibitory compositions may be active as alpha-9 integrin antagonists. That is, persons skilled in the art will recognize that compounds that are characterized as 30 inhibiting or enhancing alpha-4/beta-1 integrin binding to its ligand(s) are strong

candidates for modulating alpha-9 integrin binding to its respective ligands. Therefore, it will be a relatively routine matter, in view of the teaching of the present invention, to identify candidate compounds, for example, by conducting database searches of the patent or chemical literature for alpha-4/beta-1 (VLA-4) inhibitory compositions.

5 Exemplary methods for further testing such compounds (e.g., for specificity and selectivity of binding, as well as for relative binding affinity) are provided in Section IV below.

With the advent of automated, high throughput screening procedures and the development of a variety of forms of combinatorial chemical libraries, persons skilled in 10 the art will recognize that it will be relatively routine to identify additional alpha-9 modulatory compounds, in view of the guidance for selecting such compounds that is provided herein. For example, but not by way of limitation to the invention, random libraries are a rich source of materials. Moreover, combinatorial libraries can be produced for many types of compounds that can be synthesized in a step-by-step fashion. Such 15 compounds include peptides, beta-turn mimetics, polysaccharides, phospholipids, hormones, prostaglandins, steroids, aromatic compounds, heterocyclic compounds, benzodiazepines, oligomeric N-substituted glycines and oligocarbamates. Large combinatorial libraries of the compounds can be constructed by the encoded synthetic libraries (ESL) method described in Affymax, WO 95/12608. Affymax. WP 0306121, 20 Columbia University WO 94/08051, Pharmacopeia, WO 95/35503, and Scripps WO 95/30642 (each of which is incorporated herein by reference for all purposes). Peptide libraries can also be generated by phage display methods. See e.g., Devlin, WO 91/18980, incorporated herein by reference.

Combinatorial libraries and other compounds are initially screened by testing in 25 an alpha-9 integrin activity assay, such as one or more of the assays described herein, and a compound is selected for use in the pharmaceutical compositions and methods of the invention if it satisfies the criteria set forth, particularly in Section IV, herein.

C. Compositions including Alpha-9 Integrin Modulatory Compounds

Test compounds, preferably, but not necessarily selected as described above and 30 tested as described below are further considered for use in pharmaceutical compositions

of the present invention if they exhibit a potency in an alpha-9 integrin assay that is comparable to threshold activities determined by certain reference compounds, as discussed below. Such compositions will be suitable for further use in pharmaceutical compositions, subject to testing in an appropriate *in vivo* model appropriate to the 5 specific target disorder and subject to appropriate tests of safety for the mammalian species to be treated. Appropriate dosages to be delivered will be estimated according to standard pharmacokinetic analyses, as discussed in Section V, below. As a general guideline, an effective dosage of a compound will be that amount of compound which is effective to produce a significant biochemical effect in the target tissue, with reference to 10 the effective concentrations determined from *in vitro* or *in vivo* assays, such as are discussed below.

IV. Screening Assays for Compounds that Regulate Alpha-9 Integrin Binding

As mentioned above, a highly enriched source of compounds suitable for 15 inclusion in the pharmaceutical compositions and methods of treatment of the present invention are compounds that are identified as alpha-4/beta-1 integrin antagonists. Such compounds are identified, either with reference to the scientific and patent literature or by empirical testing in an alpha-4/beta-1 integrin binding assay, as exemplified in Part A below. Part B describes exemplary alpha-9 integrin activity assays that provide 20 information on alpha-9 integrin modulatory compounds in accordance with the present invention.

A. Alpha-4/beta-1 Integrin Binding and Activity Assays

Assays and test systems for determining whether a test compound is active in 25 binding to and modulating activity of alpha-4/beta-1 integrin are well known in the art. By way of example, but not limitation, such assays include *in vitro* assays which measure the ability of alpha-4/beta-1 integrin present on cells known to bind to one or more of its ligands, such as VCAM-1. An exemplary cell-soluble VCAM-1 protein assay suitable for this purpose is detailed in Example 1 herein, and is also described further in the parent applications, U.S. Patent Application No. 08/904,424, filed July 31, 1997, and U.S. 30 Provisional Application No. 60/054,453, filed August 1, 1997, both of which are

incorporated herein in their entirieties. These documents also describe results that identify several hundred effective alpha-4/beta-1 integrin antagonist compounds in this assay.

Briefly, in order to test compounds in this assay, compounds are synthesized, obtained from commercial sources, including a screening library, as discussed in Section 5 III, above. In experiments carried out in support of the present invention, compounds were synthesized, for example, as detailed in Example 4 herein. Test compounds are then added to the screening assay, incubated, and the amount of binding between alpha-4/beta-1 integrin and its ligand, for example, VCAM-1, is measured as detailed in Example 1.

10 By way of example, an appropriate assay for measuring this interaction employs an antibody which binds to an activation/ligand-induced epitope on the beta-1 subunit. It therefore binds only to ligand activated cells and can therefore be used as a measure of how much ligand is bound (or, conversely, displaced) in the presence of test compound.

Briefly, in experiments carried out in support of the present invention, the activity 15 of $\alpha 4\beta 1$ integrin was measured by the interaction of soluble VCAM-1 with a human T-cell line (Jurkat) which expresses high levels of $\alpha 4\beta 1$ integrin. Recombinant soluble VCAM-1 was expressed as a chimeric fusion protein containing the seven extracellular domains of VCAM-1 on the N-terminus and the human IgG1 heavy chain constant region on the C-terminus. The detector antibody, termed "15/7" was raised as a monoclonal 20 antibody against immunopurified $\alpha 4\beta 1$ integrin and was selected on the basis of surface reactivity with U937 cells (ATCC; CRL 1593), then screened for differential reactivity with Jurkat and THP-1 cells (ATCC; TIB-202). This antibody was further characterized to have the reactivity described above (Yednock). Antibodies similar to the 15/7 25 antibody have been prepared by other investigators (Luque, et al, 1996, *J. Bio. Chem.* 271:11067) and may be used in this assay.

Jurkat cells were incubated with Mn^{2+} and 15/7 antibody on ice. Mn^{2+} activates the receptor to enhance ligand binding, and 15/7 recognizes an activated/ligand occupied conformation of $\alpha 4\beta 1$ integrin and locks the molecule into this conformation thereby stabilizing the VCAM-1/ $\alpha 4\beta 1$ integrin interaction. Cells were then incubated for 30 30 minutes at room temperature with candidate compounds, in various concentrations using

a standard 5-point serial dilution. Soluble recombinant VCAM-1 fusion protein was then added to Jurkat cells and incubated for 30 minutes on ice. Cells were then washed two times and resuspended in PE-conjugated goat F(ab')2 anti-mouse IgG Fc (Immunotech, Westbrook, ME) incubated on ice, in the dark, for 30 minutes. Cells were washed twice and analyzed with a standard fluorescence activated cell sorter ("FACS") analysis as described in Yednock, et al., *supra*. Compounds having an IC₅₀ of less than about 1mM, and preferably less than about 100 μM possess sufficient binding activity to be considered for further testing.

Table 1 lists exemplary alpha-4/beta-1 integrin inhibitory compounds that were found to have activity in the foregoing assay. Moreover, experiments in support of the present invention revealed that all 373 compounds described in parent applications U.S. Patent Application No. 08/904,424, filed July 31, 1997, and U.S. Provisional Application No. 60/054,453, filed August 1, 1997, and/or in co-owned application PCT/US98/____ filed concurrently herewith, all of which are incorporated herein by reference, exhibited sufficient alpha-4-beta-1 integrin inhibitory activity to be considered candidates for alpha-9 screening, as described in Part B, below.

Table 1

<i>Compound</i>	<i>Name</i>
1	N-(toluene-4-sulfonyl)-L-proyl-L-4(4-methylpiperazin-1-ylcarbonyloxy)phenylalanine
2	N-(toluene-4-sulfonyl)-L-proyl-L-4(N,N-dimethylcarbamyloxy)phenylalanine
3	N-(1-methylpyrazole-4-sulfonyl)-L-proyl-L-4-(N,N-dimethylcarbamyloxy)phenylalanine
4	N-(toluene-4-sulfonyl)-L-(1,1-dioxo-5,5-dimethyl)thiaprolyl-L-4-(N,N-dimethylcarbamyloxy)phenylalanine
5	N-(toluene-4-sulfonyl)-N-methyl-L-alaninyl-L-4-(N,N-dimethylcarbamyloxy)phenylalanine
6	N-(toluene-4-sulfonyl)-L-[1,1-dioxo)thiamorpholin-3-carbonyl]-L-4-(N,N-dimethylcarbamyloxy)phenylalanine
7	N-(N-p-toluenesulfonyl)proyl-4-(piperazinoyloxy)phenylalanine
8	N-(N-p-toluenesulfonyl)sarcosyl-4-(N,N-dimethylcarbamyloxy)phenylalanine
9	N-(toluene-4-sulfonyl)-L-(5,5-dimethyl)thiaprolyl-L-4-[3-(N,N-dimethyl)propoxy]phenylalanine

It is understood that alpha-4/beta-1 integrin modulatory activity may alternatively be measured in one or more of other appropriate assays known and available to those skilled in the art. While absolute inhibitory concentration values may vary from assay to assay and operator to operator, compounds that inhibit (or enhance) activity at a concentration no higher than about 1 mM, and preferably no higher than about 100 μ M should be considered as candidates for alpha-9 integrin modulatory agents.

10 B. Alpha-9 Integrin Binding and Activity Assays

Example 2 describes exemplary assays for measuring alpha-9 integrin binding and activity. A convenient assay is one similar to the one described above, with reference to binding of alpha-4-beta-1 integrin to VCAM-1, using the same 15/7 antibody that recognizes activated beta-1 subunit, but substituting into the assay cells expressing the alpha-9 subunit. In experiments carried out in support of the present invention, SW480 cells were transfected with a plasmid containing an expressable coding region for the alpha-9 subunit, as described by Yokosaki, *et al.* (1994, 1996), both of which references are incorporated herein by reference. Compounds were assessed for ability to interfere with binding of these cells to tenascin.

20 Compounds 1-9 described herein potently induced the ligand-occupied epitope (15/7) on the α_9 transfected cells, but not in control mock-transfected cells, at concentrations less than about 1 μ M. Furthermore, activity of these compounds in a binding saturation assay was found to correspond to their abilities to inhibit α_9 -dependent cell adhesion to tenascin according to the methods set forth in Example 2. Based on these experiments, 25 alpha-9 inhibitory activity can be defined as inhibition of alpha-9 integrin binding to tenascin, as evidenced by an IC_{50} or effective concentration of less than about 100 μ M, and preferably less than about 20 μ M.

30 More generally, it is appreciated that active alpha-9 integrin antagonist compounds useful in the pharmaceutical compositions and methods of the invention will have activities that are defined relative to the potencies exemplified by the nine

compounds described above. That is, in accordance with a preferred embodiment of the present invention, active alpha-9 antagonist compounds have activities reflecting at least 1/1000 and preferably at least 1/100 the potency of the lowest activity compound exemplified herein. Thus, a practitioner engaged in screening compounds will know to 5 test the above-listed compounds as reference standards and to compare the activities of test compounds to the activities of the reference standards described above. According to this preferred embodiment of the invention, a test compound will be considered active if it exhibits an activity that is at least 1/1000, and preferably at least 1/100 that of the lowest activity compound exemplified above. By way of illustration, if the lowest 10 activity reference standard compound in a given assay were to exhibit an IC_{50} of 1 μM , test compounds exhibiting IC_{50} s as high as 1000 μM (1 mM) and preferably 100 μM would be considered to be active alpha-9 antagonists in accordance with the present invention.

15 ~~Methods for preparing the above-referenced reference standard compounds are~~
found, for example in Example 4 herein (compounds 1-6), in parent application U.S. Patent Application No. 08/904,424, filed July 31, 1997, or U.S. Provisional Application No. 60/054,453, filed August 1, 1997 (both of which are concurrently filed with the present application as PCT/US98/____), or in U.S. Patent Application No. 08/920,394, filed July 31, 1997, filed concurrently as PCT/US98/____, all of which are incorporated 20 herein by reference.

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While many of the alpha-4/beta-1 integrin antagonists tested exhibited equivalent activities in the alpha-4/beta-1 integrin and alpha-9 integrin activity assays described herein, compounds were also found which 10-fold or greater selectivity for inhibition in one of the assays, as compared to the other.

All nine reference compounds described herein tested exhibited at least a 100-fold selectivity in the alpha-9 integrin assay, compared to the $\alpha_5\beta_1$ integrin and $\alpha_4\beta_1$ integrin assays and were inactive in the $\alpha L\beta_2$ integrin assay. Thus, these compounds are characterized as selective for alpha-9 integrin activity, as opposed to $\alpha_5\beta_1$, $\alpha_4\beta_1$ or $\alpha L\beta_2$ activity. Such selectivity may be desirable when specificity of activity is particularly desirable. Guided by the teachings of the present specification and the particular therapeutic application for which a particular test compound is to be used, practitioners skilled in the art will be able to (a) determine appropriate drug activity assays for purposes of comparison, and (b) select a selectivity ratio that is acceptable in the context of such therapeutic application.

D. Criteria for Selection of Alpha-9 Integrin Modulatory Compounds

Active alpha-9 integrin modulatory compounds in accordance with the present invention increase or decrease binding between alpha-9 integrin and one or more of its ligands, for example tenascin, osteopontin or VCAM-1, at a concentration that is of sufficient potency to provide a pharmaceutical composition. Generally, it is appreciated that useful drugs will be active *in vitro* or *in vivo* at concentrations less than about 100 μ M and preferably less than about 20 μ M. Therefore, in accordance with the present invention, useful alpha-9 integrin modulatory compounds will be active in this concentration range in an appropriate alpha-9 integrin activity, as described herein.

Exemplified herein are a number of active alpha-9 integrin antagonist or inhibitory compounds all of which have the requisite potency to be active. In accordance with the present invention, it is suggested that these compounds can be used as reference standards in the exemplified alpha-9 antagonist activity assay or in any other appropriate alpha-9 activity assay. A compound that is run in the same assay will be considered active if it exhibits an activity that is at least 1/1000, and preferably at least 1/100 the

potency of the lowest activity reference compound selected from compounds 1-9 illustrated herein.

According to a particularly useful embodiment of the present invention exemplified herein, it is appreciated that compounds having alpha-4/beta-1 modulatory activity form a particularly useful "library" of starting compounds for identifying alpha-9 modulatory compounds. This is a useful, but not essential criterion for selecting compounds for use in the pharmaceutical compositions and methods of the present invention.

Additionally, it is appreciated that it may be advantageous to select alpha-9 modulatory compounds that are relatively selective for modulating alpha-9 integrin activity, as compared to other integrins or other pharmacological activities. Suggested criteria for selectivity are provided above.

It is further appreciated that compounds useful in the pharmaceutical compositions and treatment methods described herein should conform to acceptable levels of toxicity; persons skilled in the art will further subject test candidate compounds in toxicity assays according to standard methods known in the art and/or mandated by the appropriate regulatory authority.

V. Utility

Alpha-9 integrin modulatory compounds selected in accordance with the present activity have utility in pharmaceutical compositions, methods of treatments. In addition, the selection assays described above are useful in identifying compounds for use in such compositions and methods.

A. Pharmaceutical Compositions and Treatment of Disorders associated with Alpha-9 Integrin Binding

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Alpha-9 integrin modulatory compounds selected in accordance with the present invention find use in a number of disorders associated with alpha-9 integrin activity. Particularly, in view of the discoveries described herein with respect to the neutrophil localization of alpha-9 integrin, as well as its ability to interact with VCAM-1, it is appreciated that alpha-9 integrin inhibitory compounds will find particular

~~utility in the treatment of a variety of disorders which include an inflammatory component, particularly those in which the inflammatory component is associated with VLA-4 binding to alpha 9 integrin~~

1. Therapeutic Indications

5 The pharmaceutical compositions of the present invention can be used to block or inhibit cellular adhesion associated with a number of diseases and disorders. For instance, a number of inflammatory disorders are associated with integrins or neutrophils. Treatable disorders include, e.g., transplantation rejection (e.g., allograft rejection), Alzheimer's disease, atherosclerosis, AIDS dementia, diabetes (including acute juvenile 10 onset diabetes), retinitis, cancer metastases, rheumatoid arthritis, various lung disorders including asthma, nephritis, and acute and chronic inflammation, including atopic dermatitis, psoriasis, myocardial ischemia, and inflammatory bowel disease (including Crohn's disease and ulcerative colitis). In preferred embodiments, the pharmaceutical compositions are used to treat inflammatory brain disorders, such as Alzheimer's disease, 15 AIDS dementia, multiple sclerosis (MS), viral meningitis and encephalitis, as well as stroke (cerebral ischemia) related disorders.

More particularly, since alpha-9 integrin binding is predictive of *in vivo* utility for inflammatory conditions mediated by alpha-9 integrin, compositions and methods of the invention can be used for treating, by way of example, airway hyper-responsiveness and 20 occlusion that occurs with chronic asthma, smooth muscle cell proliferation in atherosclerosis, vascular occlusion following angioplasty, fibrosis and glomerular scarring as a result of renal disease, aortic stenosis, hypertrophy of synovial membranes in rheumatoid arthritis, and inflammation and scarring that occur with the progression of ulcerative colitis and Crohn's disease

25 In accordance with the present invention, it is appreciated that alpha-9 integrin modulatory compounds, particularly those exhibiting inhibitory activity, will find utility in treating the many, if not all, of the foregoing disorders. Compounds selected for inhibitory activity in accordance with the methods described herein are then tested in appropriate animal models, for example, to determine dosage, volumes of distribution

and the like. Efficacy may also be confirmed in such models, which are well known in the art.

For example, appropriate in vivo models for demonstrating efficacy in treating inflammatory responses include an asthma model in mice, rats, guinea pigs, goats or primates, as well as other inflammatory models in which alpha-9 integrins are implicated.

Asthma is a disease characterized by increased responsiveness of the tracheobronchial tree to various stimuli potentiating paroxysmal constriction of the bronchial airways. The stimuli cause release of various mediators of inflammation from IgE-coated mast cells including histamine, eosinophilic and neutrophilic chemotactic factors, leukotrienes, prostaglandin and platelet activating factor. Release of these factors recruits basophils, eosinophils and neutrophils, which cause inflammatory injury. In accordance with the present invention, it is believed that airway hyper-responsiveness and occlusion that occur with chronic asthma are mediated, at least in part, by alpha-9 integrin binding interactions. This is verified in a standard model such as described in Example 3, herein.

Inflammatory bowel disease is a collective term for two similar diseases referred to as Crohn's disease and ulcerative colitis. Crohn's disease is an idiopathic, chronic ulceroconstrictive inflammatory disease characterized by sharply delimited and typically transmural involvement of all layers of the bowel wall by a granulomatous inflammatory reaction. Any segment of the gastrointestinal tract, from the mouth to the anus, may be involved, although the disease most commonly affects the terminal ileum and/or colon. Ulcerative colitis is an inflammatory response limited largely to the colonic mucosa and submucosa. Lymphocytes and macrophages are numerous in lesions of inflammatory bowel disease and may contribute to inflammatory injury

Atherosclerosis is a disease of arteries (e.g., coronary, carotid, aorta and iliac). The basic lesion, the atheroma, consists of a raised focal plaque within the intima, having a core of lipid and a covering fibrous cap. Atheromas compromise arterial blood flow and weaken affected arteries. Myocardial and cerebral infarcts are a major consequence of this disease. Macrophages and leukocytes are recruited to atheromas and contribute to inflammatory injury.

Rheumatoid arthritis is a chronic, relapsing inflammatory disease that primarily causes impairment and destruction of joints. Rheumatoid arthritis usually first affects the small joints of the hands and feet but then may involve the wrists, elbows, ankles and knees. The arthritis results from interaction of synovial cells with leukocytes that 5 infiltrate from the circulation into the synovial lining of the joints. See e.g., Paul, Immunology (3d ed., Raven Press, 1993).

Another indication for the compounds of this invention is in treatment of organ or graft rejection mediated by VLA-4. Over recent years there has been a considerable improvement in the efficiency of surgical techniques for transplanting tissues and organs 10 such as skin, kidney, liver, heart, lung, pancreas and bone marrow. Perhaps the principal outstanding problem is the lack of satisfactory agents for inducing immunotolerance in the recipient to the transplanted allograft or organ. When allogeneic cells or organs are transplanted into a host (i.e., the donor and donee are different individuals from the same species), the host immune system is likely to mount an immune response to foreign 15 antigens in the transplant (host-versus-graft disease) leading to destruction of the transplanted tissue. CD8+ cells, CD4 cells and monocytes are all involved in the rejection of transplant tissues. Compounds of this invention which bind to alpha-9 integrin are useful, *inter alia*, to block alloantigen-induced immune responses in the donee thereby preventing such cells from participating in the destruction of the transplanted 20 tissue or organ. See, e.g., Paul et al., Transplant International 9, 420-425 (1996); Georcynski et al., Immunology 87, 573-580 (1996); Georcynski et al., Transplant. Immunol. 3, 55-61 (1995); Yang et al., Transplantation 60, 71-76 (1995); Anderson et al., APMIS 102, 23-27 (1994).

A related use for compounds of this invention which bind to alpha-9 integrin is in 25 modulating the immune response involved in "graft versus host" disease (GVHD). See e.g., Schlegel et al., J. Immunol. 155, 3856-3865 (1995). GVHD is a potentially fatal disease that occurs when immunologically competent cells are transferred to an allogeneic recipient. In this situation, the donor's immunocompetent cells may attack tissues in the recipient. Tissues of the skin, gut epithelia and liver are frequent targets and 30 may be destroyed during the course of GVHD. The disease presents an especially severe

problem when immune tissue is being transplanted, such as in bone marrow transplantation; but less severe GVHD has also been reported in other cases as well, including heart and liver transplants. The therapeutic agents of the present invention are used, *inter alia*, to block activation of the donor T-cells thereby interfering with their 5 ability to lyse target cells in the host.

A further use of the compounds of this invention is inhibiting tumor metastasis. Several tumor cells have been reported to express integrins, such as VLA-4 and block adhesion of such cells to endothelial cells. Steinback et al., *Urol. Res.* 23, 175-83 (1995); Orosz et al., *Int. J. Cancer* 60, 867-71 (1995); Freedman et al., *Leuk. Lymphoma* 13, 47-10 52 (1994); Okahara et al., *Cancer Res.* 54, 3233-6 (1994).

A further use of the compounds of this invention is in treating multiple sclerosis. Multiple sclerosis is a progressive neurological autoimmune disease that affects an estimated 250,000 to 350,000 people in the United States. Multiple sclerosis is thought to be the result of a specific autoimmune reaction in which certain leukocytes attack and 15 initiate the destruction of myelin, the insulating sheath covering nerve fibers. In an animal model for multiple sclerosis, murine monoclonal antibodies directed against integrins such as VLA-4 have been shown to block the adhesion of leukocytes to the endothelium, and thus prevent inflammation of the central nervous system and subsequent paralysis in the animals.

20 2. Pharmaceutical Compositions

Pharmaceutical compositions of the invention are suitable for use in a variety of drug delivery systems. Suitable formulations for use in the present invention are found in Remington's Pharmaceutical Sciences, Mace Publishing Company, Philadelphia, PA, 17th ed. (1985). Such pharmaceutical compositions are particularly useful in treating 25 diseases having an inflammatory component, such as those discussed in Part 1, above.

Pharmaceutical compositions of the present invention can also be used in *in vivo* diagnostic imaging to identify, e.g., sites of inflammation, radioisotopes are typically used in accordance with well known techniques. The radioisotopes may be bound to the peptide either directly or indirectly using intermediate functional groups. For instance, 30 chelating agents such as diethylenetriaminepentacetic acid (DTPA) and

ethylenediaminetetraacetic acid (EDTA) and similar molecules have been used to bind proteins to metallic ion radioisotopes. The complexes can also be labeled with a paramagnetic isotope for purposes of *in vivo* diagnosis, as in magnetic resonance imaging (MRI) or electron spin resonance (ESR), both of which are well known. In general, any conventional method for visualizing diagnostic images can be used. Usually gamma- and positron-emitting radioisotopes are used for camera imaging and paramagnetic isotopes are used for MRI. Thus, the compounds can be used to monitor the course of amelioration of an inflammatory response in an individual. By measuring the increase or decrease in macrophages and/or neutrophils expressing alpha-9 integrin it is possible to determine whether a particular therapeutic regimen aimed at ameliorating the disease is effective.

Compounds having the desired biological activity may be modified as necessary to provide desired properties such as improved pharmacological properties (e.g., *in vivo* stability, bio-availability), or the ability to be detected in diagnostic applications. For instance, inclusion of one or more D-amino acids in the sulfonamide compositions described herein typically increases *in vivo* stability. Stability can be assayed in a variety of ways such as by measuring the half-life of the proteins during incubation with peptidases or human plasma or serum. A number of such protein stability assays have been described (see, e.g., Verhoef, et al., *Eur. J. Drug Metab. Pharmacokinet.*, 1990, 15(2):83-93).

In order to enhance serum half-life, the compounds may be encapsulated, introduced into the lumen of liposomes, prepared as a colloid, or other conventional techniques may be employed which provide an extended serum half-life of the compounds. A variety of methods are available for preparing liposomes, as described in, e.g., Szoka, et al., U.S. Patent Nos. 4,235,871, 4,501,728 and 4,837,028 each of which is incorporated herein by reference.

Appropriate treatment dosages and dosage schedules are determined in accordance with the condition being treated, and a number of variables, including, but not limited to the intended mode of administration, the pharmacokinetics of the active

compound, the size of the subject. For example, for intravenous administration, the dose will typically be in the range of about 20 μg to about 500 μg per kilogram body weight, preferably about 100 μg to about 300 μg per kilogram body weight. Suitable dosage ranges for intranasal administration are generally about 0.1 μg to 1 mg per kilogram body weight. Dosages can be based on appropriate estimates or can be determined empirically by persons skilled in the art. Generally, relative dosages can be estimated based on comparisons of potencies in one or more of the screening assays described herein.

Effective doses can be extrapolated from dose-response curves derived from *in vitro* or animal model test systems. In general, effective dosages can be estimated from the predictive *in vitro* assays described herein. That is, an effective dose is calculated to produce at the target tissue(s) in the body, a concentration of compound that is in the range of 1/10 to 10-times the concentration of compound IC_{50} in such an assay.

The compositions administered to a patient are in the form of pharmaceutical compositions described above. These compositions may be sterilized by conventional sterilization techniques, or may be sterile filtered. The resulting aqueous solutions may be packaged for use as is, or lyophilized, the lyophilized preparation being combined with a sterile aqueous carrier prior to administration. The pH of the compound preparations typically will be between 3 and 11, more preferably from 5 to 9 and most preferably from 7 to 8. It will be understood that use of certain of the foregoing excipients, carriers, or stabilizers will result in the formation of pharmaceutical salts.

The amount administered to the patient will vary depending upon what is being administered, the purpose of the administration, such as prophylaxis or therapy, the state of the patient, the manner of administration, and the like. In therapeutic applications, compositions are administered to a patient already suffering from a disease in an amount sufficient to cure or at least partially arrest the symptoms of the disease and its complications. An amount adequate to accomplish this is defined as "therapeutically effective dose." Amounts effective for this use will depend on the disease condition being treated as well as by the judgment of the attending clinician depending upon factors such as the severity of the inflammation, the age, weight and general condition of the patient, and the like.

B. Drug Screening Assays

5 The screening assays described herein are useful in identifying new compounds for use in the treatment methods and pharmaceutical compositions described above. At its most basic, the screening assay includes testing a candidate compound for its ability to interfere with (or enhance) binding between alpha-9 integrin and one or more of its ligands, such as osteopontin, VCAM-1 or tenascin. The test compounds can also be
10 tested for the ability to competitively inhibit such binding, or between alpha-9 integrin and a labeled compound known to bind alpha-9 integrin such one of the compounds described herein or antibodies to alpha-9 integrin.

Preferably, the screening assay is used to identify alpha-9 integrin inhibitory compounds, since such compounds are, in accordance with the invention, particularly
15 useful in treating inflammation in a variety of conditions, as discussed above. According to this aspect of the invention, a test compound is added to an assay system configured to detect binding between alpha-9 integrin and one or more of its ligands, such as osteopontin, VCAM-1 or tenascin, and compounds are tested for ability to inhibit such binding. Particularly useful compounds are those which exhibit activity which is in the
20 range of, or at most about 100-times less potent than, the activity ranges defined by the reference standards provided herein (see Table 1).

As mentioned above, test compounds can be selected from a variety of sources, including combinatorial libraries, fermentation broths and the like. A particularly good source, identified herein, is the pool of compounds that are known to inhibit or are found
25 to inhibit binding between alpha-4/beta-1 integrin (VLA-4) and one or more of its ligands, such as VCAM-1.

A number of formats can be used for assays for screening for drugs. In a preferred embodiment, assays will be adapted for high throughput screening. For example, alpha-9 integrin or membranes from cells expressing alpha-9 integrin can be
30 immobilized on a solid surface, such as a microtiter plate or glass fiber filter optionally

adapted with binding aids such as antibodies to a non-ligand binding portion of the molecule. Such assay formats generally employ at least one detectably-labeled assay components. The labeling systems can be in a variety of forms. The label may be coupled directly or indirectly to the desired component of the assay according to methods 5 well known in the art. A wide variety of labels may be used. The component may be labeled by any one of several methods. The most common method of detection is the use of autoradiography with ^3H , ^{125}I , ^{35}S , ^{14}C , or ^{32}P labeled compounds and the like. Non-radioactive labels include ligands which bind to labeled antibodies, fluorophores, chemiluminescent agents, enzymes and antibodies which can serve as specific binding 10 pair members for a labelled ligand. The choice of label depends on sensitivity required, ease of conjugation with the compound, stability requirements, and available instrumentation.

15 *In vitro* uses of pharmaceutical compositions of the invention include diagnostic applications such as monitoring inflammatory responses by detecting the presence of macrophages, including neutrophils, expressing alpha-9 integrin. Compositions of this invention can also be used for isolating or labeling such cells.

20 For assays to measure the ability to block adhesion to brain endothelial cells, the assays described in International Patent Application Publication No. WO 91/05038 are particularly preferred, as adapted to the reagents described herein, according to methods well within the skill of the practitioner. This application is incorporated herein by reference in its entirety.

The following examples illustrate, but in no way are intended to limit the present invention.

25

Examples

Example 1: Binding of Compounds to $\alpha_4\beta_1$ Integrin (VLA-4)

A. 15/7 Antibody Assay

The cell adhesion assay described below is based on an assay detailed in a publication by Yednock, *et al.* (1995), which is incorporated herein by reference. An in vitro assay was used to assess binding of candidate compounds to $\alpha_4\beta_1$ integrin. Compounds which bind in this assay can be used to assess VCAM-1 levels in biological samples by conventional assays (e.g., competitive binding assays). This assay is sensitive to IC₅₀ values as low as about 1 nM.

The activity of $\alpha_4\beta_1$ integrin was measured by the interaction of soluble VCAM-1 with Jurkat cells (e.g., American Type Culture Collection Nos. TIB 152, TIB 153, and CRL 8163; American Type Culture Collection, Manassas, VA), a human T-cell line which expresses high levels of $\alpha_4\beta_1$ integrin. VCAM-1 interacts with the cell surface in an $\alpha_4\beta_1$ integrin-dependent fashion (Yednock).

Recombinant soluble VCAM-1 was expressed as a chimeric fusion protein containing the seven extracellular domains of VCAM-1 on the N-terminus and the human IgG1 heavy chain constant region on the C-terminus. The VCAM-1 fusion protein was made and purified by the manner described by Yednock, *supra*. Jurkat cells were grown in RPMI 1640 supplemented with 10% fetal bovine serum, penicillin, streptomycin and glutamine as described by Yednock, *supra*. Jurkat cells were incubated with 1.5 mM MnCl₂ and 5 μ g/mL 15/7 antibody for 30 minutes on ice. Mn²⁺ activates the receptor to enhance ligand binding, and 15/7 is a monoclonal antibody that recognizes an activated/ligand occupied conformation of $\alpha_4\beta_1$ integrin and locks the molecule into this conformation thereby stabilizing the VCAM-1/ $\alpha_4\beta_1$ integrin interaction. Yednock, *et al.*, *supra*. Antibodies similar to the 15/7 antibody have been prepared by other investigators (Luque, *et al.*, 1996, *J. Bio. Chem.* 271:11067) and may be used in this assay.

Cells were then incubated for 30 minutes at room temperature with candidate compounds, in various concentrations ranging from 66 μ g/mL to 0.01 μ g/mL using a standard 5-point serial dilution. 15 μ L soluble recombinant VCAM-1 fusion protein was then added to Jurkat cells and incubated for 30 minutes on ice. (Yednock *et al.*, *supra*). Cells were then washed two times and resuspended in PE-conjugated goat F(ab')2 anti-mouse IgG Fc (Immunotech, Westbrook, ME) at 1:200 and incubated on ice, in the dark,

for 30 minutes. Cells were washed twice and analyzed with a standard fluorescence activated cell sorter ("FACS") analysis as described in Yednock, et al., *supra*.

When tested in this assay, each of the compounds as described in Examples 1-373 of parent application USSN 08/904,424 (PCT/US98/_____) or the corresponding carboxylic acids of the ester compounds, i.e. the prodrugs) exhibited IC₅₀s of 15 μ M or less.

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B. In vitro Saturation Assay For Determining Binding of Candidate Compounds to $\alpha_4\beta_1$

Log-growth Jurkat cells were washed and resuspended in normal animal plasma containing 20 μ g/ml of the 15/7 antibody (described in the above example).

The Jurkat cells were diluted two-fold into either normal plasma samples containing known candidate compound amounts in various concentrations ranging from 66 μ g/mL to 0.01 μ g/mL, using a standard 12 point serial dilution for a standard curve, or into plasma samples obtained from the peripheral blood of candidate compound-treated animals.

Cells were then incubated for 30 minutes at room temperature, washed twice with phosphate-buffered saline ("PBS") containing 2% fetal bovine serum and 1mM each of calcium chloride and magnesium chloride (assay medium) to remove unbound 15/7 antibody. The cells were then exposed to phycoerythrin-conjugated goat F(ab')2 anti-mouse IgG Fc (Immunotech, Westbrook, ME), which has been adsorbed for any non-specific cross-reactivity by co-incubation with 5% serum from the animal species being studied, at 1:200 and incubated in the dark at 4°C for 30 minutes.

Cells were washed twice with assay medium and resuspended in the same. They are then analyzed with a standard fluorescence activated cell sorter ("FACS") analysis as described in Yednock et al. *J. Bio. Chem.*, 1995, 270:28740.

The data were graphed as fluorescence versus dose, e.g., in a normal dose-response fashion. The dose levels that result in the upper plateau of the curve represent the levels needed to obtain efficacy in an in vivo model.

This assay may also be used to determine the plasma levels needed to saturate the binding sites of other integrins, such as the $\alpha_9\beta_1$ integrin, using appropriate cells expressing alpha-9 integrin, as described in Example 2, below.

Example 2: Cell Adhesion to Tenascin

The following assay was first described by Yokosaki, *et al.* (1994) and can be used to estimate serum levels of compounds required for treating inflammatory diseases 5 related to alpha-9 integrin binding, as exemplified by the asthma model detailed in Example 3, below.

Evaluation of cell attachment to extracellular matrix proteins was performed as follows: Briefly, wells of non-tissue culture-treated polystyrene 96-well flat- bottom microtiter plates (Linbro/Titertek, Flow Laboratories, McLean, VA) were coated by 10 incubation with intact tenascin (10 μ g/ml) or recombinant tenascin fragments (1 μ g/ml to 10 μ g/ml) in PBS at 37 °C for 1 h or at 4 °C for 16 h. Wells were washed with PBS and then blocked with 1% bovine serum albumin in DMEM. 50,000 cells (SW480 cell line 15 transfected with alpha-9 integrin, (Yokosaki, 1996) a gift of D. Sheppard, Dept. of Medicine, University of California, San Francisco) were added to each well in 200 μ l of serum-free DMEM containing 0.05% bovine serum albumin. For blocking experiments, 20 cells were incubated with the relevant reagent for 30 min at 4 °C before plating. Plates were centrifugated at 10 x g for 5 min and then incubated for 1 h at 37 °C in a humidified atmosphere with 5% CO₂. Nonadherent cells were removed by centrifugation top-side-down at 48 x g for 5 min. The attached cells were fixed with 1% formaldehyde and stained with 0.5 crystal violet, and excess dye was washed off with PBS. The cells were solubilized in 50 μ l of 2% Triton X-100 and quantified by measuring the absorbance at 595 nm in a Microplate Reader (Bio-Rad). Positive adhesion results in this assay correlate with increased absorbance.

Isolated populations of neutrophils can also be used in the foregoing assay in 25 place of the transfected cells. In addition, ligands such as fibronectin, VCAM-1 or osteopontin can be substituted in this assay.

Example 3: Asthma Model

Inflammatory conditions mediated by alpha-9 integrin include, for example, airway hyper-responsiveness and occlusion that occurs with chronic asthma. The

following describes an asthma model which can be used to study the *in vivo* effects of the compounds of this invention for use in treating asthma.

Following the procedures described by Abraham et al, *J. Clin. Invest.*, 93:776-787 (1994) and Abraham et al, *Am J. Respir Crit Care Med*, 156:696-703 (1997), both of which are incorporated by reference in their entirety, compounds of this invention are formulated into an aerosol and administered to sheep which are hypersensitive to *Ascaris suum* antigen. Compounds which decrease the early antigen-induced bronchial response and/or block the late-phase airway response, e.g., have a protective effect against antigen-induced late responses and airway hyper-responsiveness ("AHR"), are considered to be active in this model. Allergic sheep which are shown to develop both early and late bronchial responses to inhaled *Ascaris suum* antigen are used to study the airway effects of the candidate compounds. Following topical anesthesia of the nasal passages with 2% lidocaine, a balloon catheter is advanced through one nostril into the lower esophagus. The animals are then intubated with a cuffed endotracheal tube through the other nostril with a flexible fiberoptic bronchoscope as a guide. Pleural pressure is estimated according to Abraham (1994). Aerosols (see formulation below) are generated using a disposable medical nebulizer that provides an aerosol with a mass median aerodynamic diameter of 3.2 μm as determined with an Andersen cascade impactor. The nebulizer is connected to a dosimeter system consisting of a solenoid valve and a source of compressed air (20 psi). The output of the nebulizer is directed into a plastic T-piece, one end of which was connected to the inspiratory port of a piston respirator. The solenoid valve is activated for 1 second at the beginning of the inspiratory cycle of the respirator. Aerosols are delivered at VT of 500 mL and a rate of 20 breaths/minute. A 0.5% sodium bicarbonate solution only was used as a control.

To assess bronchial responsiveness, cumulative concentration-response curves to carbachol are generated according to Abraham (1994). Bronchial biopsies are taken prior to and following the initiation of treatment and 24 hours after antigen challenge. Bronchial biopsies are performed according to Abraham (1994). An *in vitro* adhesion study of alveolar macrophages is also performed according to Abraham (1994), and a percentage of adherent cells is calculated.

Aerosol Formulation

A solution of the candidate compound in 0.5% sodium bicarbonate/saline (w/v) at a concentration of 30.0 mg/mL is prepared using the following procedure:

A. Preparation of 100 mL of 0.5% Sodium Bicarbonate / Saline Stock Solution:

- 5 1. Add 0.5 g sodium bicarbonate into a 100 mL volumetric flask.
2. Add approximately 90.0 mL saline and sonicate until dissolved.
3. Q.S. to 100.0 mL with saline and mix thoroughly.

B. Preparation of 10.0 mL of 30.0 mg/mL Candidate Compound:

- 10 1. Add 0.300 g of the candidate compound into a 10.0 mL volumetric flask.
2. Add approximately 9.7 mL of 0.5% sodium bicarbonate/saline stock solution.
3. Sonicate until the candidate compound is completely dissolved.
4. Q.S. to 10.0 mL with 0.5% sodium bicarbonate/saline stock solution and mix thoroughly.

15 15 Example 4: Synthesis of alpha-9 Integrin-modulatory Compounds

This Example provides representative organic syntheses of pre-cursors and reference standard compounds described herein. Parent applications U.S. Patent

Application No. 08/904,424, filed July 31, 1997, and U.S. Provisional Application No. 60/054,453, filed August 1, 1997, as well as co-owned, concurrently filed US Patent

20 Application No. _____ and corresponding PCT application PCT/US98/_____
all of which applications are incorporated herein by reference in their entireties, are

referenced specifically for their methods of preparation of these and many other related compounds.

A. General Reference Methods

25 Method 1: N-Tosylation Procedure

N-Tosylation of the appropriate amino acid was conducted via the method of Cupps, Boutin and Rapoport J. Org. Chem. 1985, 50, 3972.

Method 2: Methyl Ester Preparation Procedure

Amino acid methyl esters were prepared using the method of Brenner and Huber Helv.

30 Chim. Acta 1953, 36, 1109.

Method 3: BOP Coupling Procedure

The desired dipeptide ester was prepared by the reaction of a suitable N-protected amino acid (1 equivalent) with the appropriate amino acid ester or amino acid ester hydrochloride (1 equivalent), benzotriazol-1-yloxy-tris(dimethylamino)phosphonium

5 hexafluorophosphate [BOP] (2.0 equivalent), triethylamine (1.1 equivalent), and DMF.

The reaction mixture was stirred at room temperature overnight. The crude product is purified flash chromatography to afford the dipeptide ester.

Method 4: Hydrogenation Procedure I

Hydrogenation was performed using 10% palladium on carbon (10% by weight) in

10 methanol at 30 psi overnight. The mixture was filtered through a pad of Celite and the filtrate concentrated to yield the desired amino compound.

Method 5: Hydrolysis Procedure I

To a chilled (0°C) THF/H₂O solution (2:1, 5 - 10 mL) of the appropriate ester was added

LiOH (or NaOH) (0.95 equivalents). The temperature was maintained at 0°C and the

15 reaction was complete in 1-3 hours. The reaction mixture was extracted with ethyl acetate and the aqueous phase was lyophilized resulting in the desired carboxylate salt.

Method 6: Ester Hydrolysis Procedure II

To a chilled (0°C) THF/H₂O solution (2:1, 5 - 10 mL) of the appropriate ester was added

LiOH (1.1 equivalents). The temperature was maintained at 0°C and the reaction was

20 complete in 1-3 hours. The reaction mixture was concentrated and the residue was taken up into H₂O and the pH adjusted to 2-3 with aqueous HCl. The product was extracted with ethyl acetate and the combined organic phase was washed with brine, dried over MgSO₄, filtered and concentrated to yield the desired acid.

Method 7: Ester Hydrolysis Procedure III

25 The appropriate ester was dissolved in dioxane/H₂O (1:1) and 0.9 equivalents of 0.5 N NaOH was added. The reaction was stirred for 3-16 hours and then concentrated. The resulting residue was dissolved in H₂O and extracted with ethyl acetate. The aqueous phase was lyophilized to yield the desired carboxylate sodium salt.

Method 8: Sulfonylation Procedure I

To the appropriately protected aminophenylalanine analog (11.2 mmol), dissolved in methylene chloride (25ml) and cooled to -78°C was added the desired sulfonyl chloride (12 mmol) followed by dropwise addition of pyridine (2 mL). The solution was allowed to warm to room temperature and was stirred for 48 hr. The reaction solution was 5 transferred to a 250 mL separatory funnel with methylene chloride (100 mL) and extracted with 1N HCl (50 mL x 3), brine (50 mL), and water (100 mL). The organic phase was dried (MgSO₄) and the solvent concentrated to yield the desired product.

Method 9: Reductive Amination Procedure

Reductive amination of Tos-Pro-p-NH₂-Phe with the appropriate aldehyde was 10 conducted using acetic acid, sodium triacetoxyborohydride, methylene chloride and the combined mixture was stirred at room temperature overnight. The crude product was purified by flash chromatography.

Method 10: BOC Removal Procedure

Anhydrous hydrochloride (HCl) gas was bubbled through a methanolic solution of the 15 appropriate Boc-amino acid ester at 0°C for 15 minutes and the reaction mixture was stirred for three hours. The solution was concentrated to a syrup and dissolved in Et₂O and reconcentrated. This procedure was repeated and the resulting solid was placed under high vacuum overnight.

Method 11: tert-Butyl Ester Hydrolysis Procedure I

20 The tert-butyl ester was dissolved in CH₂Cl₂ and treated with TFA. The reaction was complete in 1-3 hr at which time the reaction mixture was concentrated and the residue dissolved in H₂O and lyophilized to yield the desired acid.

B. Preparations

Prep 1 Synthesis of N-(Toluene-4-sulfonyl)-L-prolyl-L-4-(4-methylpiperazin-1-ylcarbonyloxy)phenylalanine Ethyl Ester

The title compound was prepared following the procedure outlined for the preparation of Prep 4 and substitution of appropriate starting materials.

NMR data were as follows:

1H NMR (CD₃)₂SO: δ = 8.33 (d, 1H), 7.70 (d, 2H), 7.41 (d, 2H), 7.24 (d, 2H), 7.00 (d, 2H), 4.52-4.44 (m, 1H), 4.09-4.00 (m, 3H), 3.53 (bs, 2H), 3.38-3.31 (m, 3H), 3.11-3.01

(m, 3H), 2.39 (s, 3H), 2.32 (bs, 4H), 2.19 (s, 3H), 1.61-1.50 (m, 3H), 1.43-1.38 (m, 1H), 1.13 (t, 3H). ^{13}C NMR (CD3)2SO): δ = 171.1, 171.1, 153.9, 149.8, 143.6, 134.1, 133.9, 130.0, 129.8, 127.4, 121.5, 61.2, 60.7, 54.2, 54.1, 53.3, 49.0, 45.7, 44.0, 43.4, 35.8, 30.5, 23.8, 21.0, 14.0.

5 **Prep 2** Synthesis of N-(Toluene-4-sulfonyl)-L-prolyl-L-4-(N,N-dimethylcarbamyl)oxyphenylalanine Ethyl Ester (Compound 2)

Into a reaction vial were combined 7.00 g (15.2 mmol, 1.0 eq) Ts-Pro-Tyr(H)-OEt and 1.86 g (15.2 mmol, 1.0 eq) DMAP. Methylene chloride (50 mL), triethylamine (2.12 mL -- 1.54 g, 15.2 mmol, 1.0 eq), and dimethylcarbamyl chloride (1.68 mL -- 1.96 g, 18.2 mmol, 1.2 eq) were then added. The vial was capped tightly, and the reaction solution swirled to obtain a homogeneous solution. The reaction solution was then heated to 40°C. After 48 h, TLC of the resulting colorless solution indicated complete conversion. The workup of the reaction solution was as follows: add 50 mL EtOAc and 50 mL hexanes to the reaction mixture, and wash with 3 x 50 mL 0.5 mL hexanes to the reaction mixture, and wash with 3 x 50 mL 0.5 M citric acid, 2 x 50 mL water, 2 x 50 mL 10% K2CO3, and 1 x 50 mL sat. NaCl. Dry with MgSO4. Filter. Evaporate to obtain 8.00 g (99%) of the title compound as a clear oil, which solidifies upon standing. Recrystallize from 5:3:2 heptane/EtOAc/CH2Cl2.

NMR data were as follows:

20 ^1H NMR (CD3)2SO): δ = 8.32 (d, 1H), 7.70 (d, 2H), 7.41 (d, 2H), 7.23 (d, 2H), 7.00 (d, 2H), 4.52-4.44 (m, 1H), 4.09-4.02 (m, 3H), 3.37-3.31 (m, 1H), 3.11-2.96 (m, 3H), 3.00 (s, 3H), 2.87 (s, 3H), 2.39 (s, 3H), 1.61-1.50 (m, 3H), 1.43-1.38 (m, 1H), 1.13 (t, 3H).

^{13}C NMR (CD3)2SO): δ = 171.1, 171.1, 154.0, 150.0, 143.6, 133.9, 133.9, 130.0, 129.8, 127.4, 121.5, 61.2, 60.6, 53.3, 49.0, 36.3, 36.1, 35.8, 30.5, 23.8, 21.0, 14.0.

25 **Prep 3:** Synthesis of N-(Toluene-4-sulfonyl)-L-prolyl-L-4-(4-methylpiperazin-1-ylcarbonyloxy)phenylalanine Isopropyl Ester

The title compound was prepared following the procedure outlined for the preparation of Prep 4 and substitution of appropriate starting materials.

NMR data were as follows:

1H NMR (CDCl₃): δ = 7.72 (d, 2H), 7.36 (d, 1H), 7.33 (d, 2H), 7.16 (d, 2H), 7.03 (d, 2H), 5.07 (Sept., 1H), 4.78 (dt, 1H), 4.08-4.05 (m, 1H), 3.67 (bs, 2H), 3.57 (bs, 2H), 3.41-3.35 (m, 1H), 3.24 (dd, 1H), 3.15-3.07 (m, 1H), 3.04 (dd, 1H), 3.46-2.43 (m, 7H), 2.34 (s, 3H), 2.05-2.02 (m, 1H). 13C NMR (CDCl₃): δ = 170.9, 170.4, 153.6, 150.5, 144.3, 5 133.2, 133.1, 130.2, 130.0, 127.9, 121.7, 69.5, 62.2, 54.7, 53.4, 49.6, 46.1, 44.3, 43.7, 37.2, 29.7, 24.1, 21.6, 21.6, 21.4.

Prep 4 Synthesis of N-(Toluene-4-sulfonyl)-L-prolyl-L-4-(4-methylpiperazin-1-ylcarbonyloxy)phenylalanine tert-Butyl Ester

Combine 41.2 g (84.34 mmol, 1.0 eq) Ts-Pro-Tyr(H)-OtBu and 17.0 g (84.34 mmol, 1.0 eq) 4-nitrophenyl chloroformate. Add 700 mL CH₂Cl₂. Cap with a septum. Attach a N₂ line. Immerse the flask in a 4:1 water/EtOH + dry ice slurry, and stir to cool to -15°C. Add 29.38 mL (21.33 g, 210.81 mmol, 2.5 eq) Et₃N over five minutes with stirring. Stir at -10 to -15°C for 1 h. Add 9.35 mL (8.45 g, 84.34 mmol, 1.0 eq) N-methyl piperazine over 3 minutes with stirring. Stir overnight while warming to room temperature. Dilute 10 with 700 mL hexanes. Wash repeatedly with 10% K₂CO₃, until no yellow color (4-nitrophenol) is seen in the aqueous layer. Wash with sat. NaCl. Dry over anhydrous MgSO₄. Filter. Evaporate. Dissolve in 500 mL EtOH, and evaporate, to remove Et₃N. Repeat once. Dissolve in 400 mL EtOH, and add 600 mL water with stirring, to precipitate a solid or oil. If an oil, stir vigorously to solidify. Isolate the solid by 15 filtration. Repeat dissolution, precipitation, and filtration, once. Rinse with water to remove traces of yellow color. High vacuum to constant mass yields the title compound as a white solid.

NMR data were as follows:

1H NMR (CDCl₃): δ = 7.72 (d, 2H), 7.33 (d, 3H), 7.17 (d, 2H), 7.02 (d, 2H), 4.71 (q, 25 1H), 4.09-4.06 (m, 1H), 3.67 (bs, 2H), 3.57 (bs, 2H), 3.41-3.34 (m, 1H), 3.22 (dd, 1H), 3.16-3.09 (m, 1H), 3.03 (dd, 1H), 2.46-2.43 (m, 7H), 2.34 (s, 3H), 2.05-2.02 (m, 1H), 1.57-1.43 (m, 3H), 1.47 (s, 9H). 13C NMR (CDCl₃): δ = 171.8, 169.9, 153.6, 150.4, 144.3, 133.4, 133.1, 130.3, 130.0, 127.9, 121.6, 82.6, 62.3, 54.5, 53.8, 49.6, 46.1, 44.3, 43.7, 37.3, 29.7, 27.8, 24.1, 21.4.

Prep 5 Synthesis of N-(Toluene-4-sulfonyl)-L-prolyl-L-4-(4-methylpiperazin-1-ylcarbonyloxy)phenylalanine (Reference Compound 1)

The title compound was prepared from the product of Prep 1 using the procedure described in Method 7.

5 NMR data were as follows:

1H NMR (CD3OD): δ = 7.74 (d, 2H), 7.42 (d, 2H), 7.26 (d, 2H), 7.04 (d, 2H), 4.58-4.54 (m, 1H), 4.16-4.12 (m, 1H), 3.70 (bs, 2H) 3.53 (bs, 2H), 3.43-3.31 (m, 1H), 3.26-3.13 (m, 7H), 2.82 (s, 3H), 2.43 (s, 3H), 1.98-1.94 (m, 1H), 1.76-1.51 (m, 3H).

13C NMR (CD3OD): δ = 175.7, 173.6, 154.8, 151.6, 146.1, 136.3, 134.8, 131.9, 131.3,

10 129.1, 122.7, 63.6, 55.9, 53.9, 50.7, 43.5, 37.6, 31.3, 25.5, 21.5.

Prep 9 Synthesis of N-(Toluene-4-sulfonyl)-L-prolyl-L-4- (N,N-dimethylcarbamyloxy)phenylalanine

The title compound was prepared from the product of Prep 2 using the procedure

15 described in Method 7.

NMR data were as follows:

1H NMR (CD3)2SO: δ = 8.13 (d, 1H), 7.70 (d, 2H), 7.41 (d, 2H), 7.23 (d, 2H), 6.99 (d, 2H), 4.51-4.44 (m, 1H), 4.11-4.09 (m, 1H), 3.40-3.34 (m, 2H), 3.11-2.94 (m, 3H), 3.00 (s, 3H), 2.87 (s, 3H), 2.39 (s, 3H), 1.59-1.36 (m, 4H). 13C NMR (CD3)2SO: δ = 172.7,

20 171.2, 153.6, 150.2, 143.8, 134.3, 134.0, 130.2, 130.0, 127.6, 121.6, 61.3, 53.2, 49.0, 36.3, 36.1, 35.9, 30.4, 23.8, 21.0.

Prep 10: Synthesis of N-(Toluene-4-sulfonyl)-L-prolyl-L-3-(N,N-dimethylcarbamyloxy)phenylalanine Ethyl Ester

25 The title compound was prepared following the procedure outlined for the preparation of Prep 2 and substitution of appropriate starting materials.

NMR data were as follows:

1H NMR (CDCl3): δ = 7.74 (m, 2H), 7.70-7.36 (m, 4H), 7.24-7.14 (m, 3H), 6.93-4.90 (m, 1H), 4.78-4.27 (m, 3H), 4.05-3.55 (m, .5H), 3.48-3.43 (m, .5H), 3.37-3.30 (m, 3H),

3.02-3.08 (bs, 3H), 2.99 (bs, 3H), 2.45 (s, 1.5H), 2.43 (s, 1.5H), 2.12 (m, 1H), 198, 1.80 (m, .5M), 1.62-1.44 (m, 2.5H), 1.29 (t, 1.5H), 1.24 (t, 1.5H).

¹³C NMR (CDCl₃): δ = 171.1, 171.0, 170.9, 154.9, 154.8, 151.8, 151.6, 144.4, 144.3, 137.6, 137.1, 133.1, 132.9, 130.0, 129.9, 129.5, 129.2, 127.9, 127.9, 126.5, 126.1, 122.9,

5 122.7, 120.7, 120.5.

Prep 11: Synthesis of N-(Toluene-4-sulfonyl)-L-(5,5-dimethyl)thiaprolyl-

L-4-(N,N-dimethylcarbamyl)oxyphenylalanine Isopropyl Ester

The title compound was prepared following the procedure outlined for the preparation of

10 Prep 2 and substitution of appropriate starting materials.

NMR data were as follows:

¹H NMR (CDCl₃): δ = 7.76 (d, 2H), 7.35 (d, 2H), 7.22 (d, 2H), 7.01 (m, 3H), 5.05 (m, 1H), 4.85 (m, 1H), 4.57 (d, 1H), 4.38 (d, 1H), 3.86 (s, 1H), 3.19-3.00 (m, 2H), 3.09 (s, 3H), 3.01 (s, 3H), 2.45 (s, 3H), 1.24 (t, 6H), 1.16 (s, 3H), 1.09 (s, 3H).

15 ¹³C NMR (CDCl₃): δ = 170.3, 168.4, 154.9, 150.6, 144.8, 132.9, 132.8, 130.3, 130.0, 128.2, 121.7, 73.4, 69.5, 54.5, 53.2, 50.4, 37.7, 36.5, 36.3, 29.0, 23.8, 21.5, 21.4.

Prep 12: Synthesis of N-(Toluene-4-sulfonyl)-L-(5,5-dimethyl)thiaprolyl-

L-4-(N,N-dimethylcarbamyl)oxyphenylalanine tert-Butyl Ester

20 The title compound was prepared following the procedure outlined for the preparation of Prep 2 and substitution of appropriate starting materials.

NMR data were as follows:

¹H NMR (CDCl₃): δ = 7.75 (d, 2H), 7.34 (d, 2H), 7.23 (d, 2H), 7.05-6.98 (m, 3H), 4.76 (m, 1H), 4.56 (d, 1H), 4.40 (d, 1H), 3.85 (s, 1H), 3.09-3.00 (m, 8H), 2.44 (s, 3H), 1.43 (s, 3H), 1.16 (s, 3H), 1.09 (s, 3H).

25 ¹³C NMR (CDCl₃): δ = 169.8, 168.3, 154.9, 150.6, 144.8, 133.2, 132.9, 130.4, 130.0, 128.2, 121.6, 82.6, 73.4, 54.6, 53.8, 50.4, 37.8, 36.5, 36.3, 29.0, 27.7, 23.8, 21.5.

Prep 13: Synthesis of N-(Toluene-4-sulfonyl)-L-(5,5-dimethyl)thiaprolyl-

L-4-(N,N-dimethylcarbamyloxy)phenylalanine

The title compound was prepared from the product of Prep 11 using the procedure described in Method 7.

NMR data were as follows:

5 ^1H NMR (CDCl₃): δ = 7.76 (d, 2H), 7.35 (d, 2H), 7.25 (d, 2H), 7.14 (d, 1H), 7.02 (d, 2H), 5.17 (br s, 1H), 4.89 (m, 1H), 4.56 (d, 1H), 4.40 (d, 1H), 3.90 (s, 1H), 3.30-3.00 (m, 8H), 2.43 (s, 3H), 1.09 (s, 6H).

13C NMR (CDCl₃): δ = 172.7, 169.3, 155.2, 150.6, 144.9, 133.1, 132.7, 130.5, 130.1, 128.1, 121.9, 73.3, 54.5, 53.3, 50.5, 36.9, 36.6, 36.4, 29.0, 23.7, 21.5.

10 Prep 18 Synthesis of N-(Toluene-4-sulfonyl)sarcosyl-L-4-(N,N-dimethylcarbamyloxy)phenylalanine Isopropyl Ester

The title compound was prepared following the procedure outlined for the preparation of Prep 2 and substitution of appropriate starting materials.

NMR data were as follows:

15 ^1H NMR (CDCl₃): δ = 7.66 (d, 2H), 7.34 (d, 2H), 7.18 (d, 2H), 7.07 (d, 2H), 6.98 (d, 1H), 5.03 (m, 1H), 4.81 (m, 1H), 3.69 (d, 1H), 3.49 (d, 1H), 3.08 (m, 2H), 3.04 (s, 3H), 2.99 (s, 3H), 2.63 (s, 3H), 2.43 (s, 3H).

13C NMR (CDCl₃): δ = 167.4, 154.9, 150.8, 144.4, 132.6, 130.2, 130.1, 127.7, 122.0, 110.9, 69.5, 57.3, 53.9, 53.0, 37.1, 36.6, 21.6, 21.4.

20 Prep 19 Synthesis of N-(Toluene-4-sulfonyl)sarcosyl-L-4-(N,N-dimethylcarbamyloxy)phenylalanine tert-Butyl Ester

The title compound was prepared following the procedure for the preparation of Prep 2 and substitution of appropriate starting materials.

25 NMR data were as follows:

^1H NMR (CDCl₃): δ = 7.67 (d, 2H), 7.34 (d, 2H), 7.19 (d, 2H), 7.03 (d, 2H), 6.98 (d, 1H), 4.76 (m, 1H), 3.67 (q, 1H), 3.06 (m, 2H), 3.16 (s, 3H), 2.99 (s, 3H), 2.64 (s, 3H), 2.43 (s, 3H), 1.42 (s, 9H).

13C NMR (CDCl3): δ = 170.0, 137.2, 154.9, 150.7, 144.3, 133.2, 132.9, 130.3, 130.0, 127.7, 121.9, 82.6, 83.9, 53.3, 37.2, 36.6, 36.4, 27.9, 21.4.

Prep 20 Synthesis of N-(Toluene-4-sulfonyl)sarcosyl-L-4-(N,N-

5 dimethylcarbamyl)oxy)phenylalanine (Reference Compound 8)

The title compound was prepared from the product of Prep 18 using the procedure described in Method 7.

NMR data were as follows:

1H NMR (CDCl3): δ = 7.41 (d, 2H), 7.10 (d, 2H), 6.98 (d, 2H), 6.75 (d, 2H), 4.42 (m,

10 1H), 3.43 (m, 2H), 3.04 (m, 2H), 2.80 (s, 3H), 2.69 (s, 3H), 2.33 (s, 3H), 2.14 (s, 3H).

13C NMR (CDCl3): δ = 174.2, 170.2, 156.9, 151.9, 145.6, 135.5, 135.2, 131.4, 131.1,

128.9, 123.0, 54.6, 54.0, 37.4, 36.8, 36.7, 21.4.

Prep 29 Synthesis of N-(Toluene-4-sulfonyl)-L-(1,1-dioxo-5,5-dimethyl)-

15 thiaprolyl-L-4-(N,N-dimethylcarbamyl)oxy)phenylalanine
tert-Butyl Ester

The product of Prep 12 was oxidized by the method of Larsson and Carlson (Acta Chemica Scan. 1994, 48, 517-525), yielding the title compound as a white solid.

NMR data were as follows:

20 1H NMR (CDCl3): δ = 7.73 (d, 2H), 7.36 (d, 2H), 7.21 (d, 2H), 7.06-6.95 (m, 3H), 4.79 (m, 1H), 4.38 (dd, 2H), 4.10 (s, 1H), 3.18-2.95 (m, 8H), 2.43 (s, 3H), 1.45 (s, 9H), 1.33 (s, 3H), 1.08 (s, 3H).

13C NMR (CDCl3): δ = 169.8, 166.2, 154.9, 120.7, 145.8, 133.0, 131.9, 130.2, 128.5, 121.9, 82.9, 68.0, 60.9, 59.3, 53.9, 37.5, 36.6, 36.3, 27.7, 21.6, 19.3, 18.5.

25

Prep 30 Synthesis of N-(1-Methylimidazolyl-4-sulfonyl)-L-prolyl-
L-4-(N,N-dimethylcarbamyl)oxy)phenylalanine

The title compound was prepared from the product of Example 106 using the procedure described in Method 11.

NMR data were as follows:

1H NMR (CDCl₃): δ = 8.07 (d, 1H), 7.75 (s, 1H), 7.71 (s, 1H), 7.25 (d, 2H), 7.01 (d, 2H), 4.71-4.66 (m, 1H), 4.28-4.24 (m, 1H), 3.77 (s, 3H), 3.42-3.05 (m, 3H), 3.09 (s, 3H), 2.96 (s, 3H), 1.84-1.69 (m, 2H), 1.61-1.54 (m, 2H).

5 13C NMR (CDCl₃): δ = 174.4, 174.1, 156.9, 151.9, 141.8, 137.7, 135.6, 131.6, 127.6, 122.9, 63.7, 54.7, 50.8, 37.4, 36.8, 36.7, 34.3, 31.6, 25.4.

Prep 31 Synthesis of N-(Toluene-4-sulfonyl)-L-(1,1-dioxo-5,5-dimethyl)-thiaprolyl-L-4-(N,N-dimethylcarbamyl)oxyphenylalanine (Reference Compound 4)

10 The title compound was prepared from the product of Prep 29 using the procedure described in Method 11.

NMR data were as follows:

1H NMR (CDCl₃): δ = 7.75 (m, 3H), 7.29 (m, 4H), 7.08 (d, 2H), 4.95 (m, 1H), 4.46-4.20 (m, 3H), 3.17 (s, 3H), 3.30-3.10 (m, 2H), 3.02 (s, 3H), 2.43 (s, 3H), 1.15 (s, 3H), 0.88 (s, 3H).

15 13C NMR (CDCl₃): δ = 127.2, 167.5, 155.8, 150.3, 145.4, 133.6, 132.6, 130.8, 130.2, 128.3, 121.9, 67.9, 65.8, 60.8, 53.9, 36.8, 36.6, 35.8, 21.6, 18.8, 15.0.

Prep 49 Synthesis of N-(Toluene-4-sulfonyl)-L-(thiamorpholin-3-carbonyl)-

20 L-4-(N,N-dimethylcarbamyl)oxyphenylalanine tert-Butyl Ester
L-Thiamorpholine-3-carboxylic acid was prepared by the method of Larsson and Carlson (Acta Chemica Scan. 1994, 48, 517-525). N-(Toluene-4-sulfonyl)-L-thiamorpholine-3-carboxylic acid was prepared using the procedure described in Method 1. The title compound was prepared following the procedure for the synthesis of Prep 2 with
25 substitution of appropriate starting materials.

NMR data were as follows:

1H NMR (CDCl₃): δ = 7.69 (d, 2H), 7.31 (d, 2H), 7.16 (d, 2H), 6.98 (d, 2H), 6.86 (d, 1H), 4.71 (m, 1H), 4.62 (m, 1H), 3.94 (m, 1H), 3.31 (m, 1H), 3.09 (m, 4H), 2.98 (s, 3H), 2.67 (m, 1H), 2.50 (m, 1H), 2.40 (s, 3H), 2.31 (m, 1H), 2.10 (m, 1H), 1.49 (s, 9H).

¹³C NMR (CDCl₃): δ= 169.9, 167.4, 154.8, 150.6, 144.2, 136.8, 132.8, 130.4, 130.2, 127.3, 121.8, 82.6, 55.2, 54.0, 43.3, 36.5, 36.3, 27.8, 25.2, 24.6, 21.4.

Prep 50 Synthesis of N-(Toluene-4-sulfonyl)sarcosyl-

5 L-4-(1,1-dioxothiomorpholin-4-ylcarbonyloxy)phenylalanine

The title compound was prepared from the product of Prep 121 using the procedure described in Method 11.

NMR data were as follows:

1H NMR (CD₃OD): δ = 7.67 (d, 2H), 7.40 (d, 2H), 7.27 (d, 2H), 7.09 (d, 2H), 4.61 (m, 1H), 4.12 (m, 2H), 3.99 (m, 2H), 3.60 (m, 2H), 3.23 (m, 8H), 2.58 (s, 3H), 2.42 (s, 3H).

10 ¹³C NMR (CD₃OD): δ= 174.2, 170.3, 155.0, 151.6, 145.6, 136.1, 135.2, 131.5, 131.1, 128.9, 123.0, 54.6, 54.0, 52.4, 52.2, 44.4, 44.0, 37.4, 36.8, 21.4.

Prep 51 Synthesis of N-(Toluene-4-sulfonyl)-L-(1,1-dioxothiamorpholin-3-carbonyl)-L-4-(N,N-dimethylcarbamyloxy)phenylalanine tert-Butyl Ester

15 The title compound was prepared from the product of Prep 49 following the procedure described by Larsson and Carlson (Acta Chemica Scan. 1994, 48, 522).

NMR data were as follows:

1H NMR (CDCl₃): δ= 7.76 (d, 2H), 7.37 (d, 2H), 7.08 (d, 2H), 6.98 (d, 2H), 6.56 (d, 1H), 4.95 (m, 1H), 4.62 (m, 1H), 3.99 (m, 2H), 3.25 (m, 1H), 3.07 (s, 3H), 2.97 (m, 8H), 2.44 (s, 3H), 1.48 (s, 9H).

20 ¹³C NMR (CDCl₃): δ= 170.0, 164.8, 154.9, 150.7, 145.4, 135.3, 132.6, 130.7, 130.3, 127.5, 122.3, 82.8, 56.1, 53.6, 49.5, 48.6, 41.6, 36.6, 36.4, 27.9, 21.6.

Prep 60 Synthesis of N-(Toluene-4-sulfonyl)-L-(1,1-dioxothiamorpholin-3-carbonyl)-L-4-(N,N-dimethylcarbamyloxy)phenylalanine (Reference Compound 6)

25 The title compound was prepared from the product of Prep 51 using the procedure described in Method 11.

NMR data were as follows:

1H NMR (CDCl₃): δ= 7.79 (d, 2H), 7.43 (d, 2H), 7.20 (d, 2H), 7.00 (d, 2H), 5.21 (m, 1H), 4.65 (m, 1H), 4.12 (m, 1H), 3.75 (m, 1H), 3.29 (m, 3H), 3.08 (s, 3H), 3.00 (m, 1H), 30 3.00 (m, 1H), 2.97 (s, 3H), 2.80 (m, 3H), 2.44 (s, 3H).

13C NMR (CDCl₃): δ= 165.1, 159.0, 147.9, 143.1, 137.6, 128.6, 126.1, 122.7, 122.6, 119.8, 114.3, 48.3, 45.8, 41.6, 34.0, 28.0, 27.8, 27.7, 12.5.

While the invention has been described with reference to specific methods and
5 embodiments, it will be appreciated that various modifications and changes may be made
without departing from the invention.